**In the Specification:**

**On page 5, the fourth full paragraph has been amended as follows:**

~~Figure 2 shows~~ Figures 2A-2B show a possible procedure for charging a miniaturized analytical system by an inventive delivery apparatus.

**On page 5, the fifth full paragraph has been amended as follows:**

~~Figure 3 shows~~ Figures 3A-3D show a delivery apparatus for miniaturized analytical systems from the prior art.

**On page 5, the sixth full paragraph has been amended as follows:**

~~Figure 4 shows~~ Figures 4A-4F show a delivery apparatus for macroscopic analytical systems from the prior art.

**The last paragraph bridging pages 8 and 9 has been amended as follows:**

~~Figure 2 shows~~ Figures 2A and 2B show a possible procedure for charging a miniaturized analytical system. The figure shows a channel system consisting of three reservoirs R1 to R3, the channel sections K1 to K4, the fluidic connections F1 to F6 and a branching point Vz. The system shown in the figure has a channel section K1 for sample delivery. The separation can be performed along channel section K2 and K3, or K2 and K4. To carry out an isotachophoretic separation, the system must be charged with a sample and appropriate buffers. In this case, the sample volume must be in contact with one buffer

(leading buffer) at one end in the direction of the separation path and with another buffer (terminating buffer) at the other end. As a result of the branching Vz of the channel system, there is the possibility of charging different leading buffers via reservoirs R2 and R3. Components which have been fractionated from the sample can be discharged via the fluidic connection F3.

**On page 9, the first full paragraph has been amended as follows:**

In order to achieve the desired arrangement of sample and buffers in the channel system, firstly, as shown diagrammatically under ~~A in the figure~~ Fig. 2A, the fluidic connections F2 (outlet), F4, F5 and F6 (inlets) are open, and the channel system is filled via the three reservoirs with the two leading buffers (via R2 and R3, shown hatched and dotted, respectively) and the terminating buffer (via R1, shown with vertical stripes). Excess buffer can exit via the fluidic connection F2. In this manner, channel section K1 fills with terminating buffer, section K3 with leading buffer (LE2) via R2, section K4 with leading buffer (LE1) via R3 and channel section K2 contains a mixture of the two leading buffers. The fluidic connections F1 and F3 remain closed during this step.

**The paragraph bridging pages 9 and 10 has been amended as follows:**

~~Part B of the figure~~ Fig. 2B shows how the sample is introduced into channel section K1 and the channel section K2 is filled with a leading buffer via R3. The fluidic connections F5 and F6 are closed and no further trailing buffer is pumped via R1 and no further leading buffer (LE2) is pumped via R2. Fluidic connection F4 is open and channel section K2 is filled with leading buffer (LE1) via R3. At the same time, fluidic connection F1 is open and the sample is fed via F1 (shown as wavy lines). Excess sample and excess leading buffer

(LE1) can exit via the open fluidic connection F2. By the leading buffer (LE1) and the sample volume being pumped simultaneously against one another, a particularly precise filling of channel sections K1 and K2 is achieved. In this manner, it is possible to perform exact charging even using pumps which have a slight pulsation.

**On page 11, the second full paragraph after the table has been amended as follows:**

Below, by way of example, some switching processes are listed for various analytical processes on an analytical unit corresponding to ~~Figure 2~~ Figures 2A and 2B:
(The voltage is applied in each case downstream of the fluidic connections)

**On page 12, the first full paragraph has been amended as follows:**

~~Figure 3 shows~~ Figures 3A-3D show a possible method for electrokinetic sample delivery in miniaturized analytical systems from the prior art. ~~Figures A, B, C and D show the~~ in individual steps of sample delivery. Figure 3A shows diagrammatically a crossed channel structure. At the ends of the channels are situated the electrodes E1 to E4. First, as shown in Figure 3B, a channel is filled with sample by applying a voltage between electrode E1 (0 V) and E2 (+500 V). Then, as shown in Figure 3C, the electrodes in the filled channel are switched to the same potential (for example E1 and E2 both at +400 V) and a voltage is applied to the separation channel system situated perpendicularly thereto (E3 = 0 V and E4 = +2.5 kV). In this manner, the sample volume which is situated at the intersection of the two channel systems is transported into the separation channel system (Figure D). The sample volume thus produced is in the range of some nanolitres or less.

**The paragraph bridging pages 12 and 13 has been amended as follows:**

~~Figure 4 shows~~ Figures 4A and 4F show a possible method for sample delivery in macroscopic analytical systems, for example the isotachophoresis instrument ItaChrom® EA 101 from I+M, Analytische Meß- und Regeltechnik, Germany. Figures A1/A2, B1/B2 and C1/C2 show the different sample delivery steps, with Figures ~~A1, B1 and C1~~ 4A, 4C and 4E showing a side view of the delivery apparatus, and Figures ~~A2, B2 and C2~~ 4B, 4D and 4Fshowing a view from above. This mechanical sample delivery apparatus consists of a stopcock K which is surrounded by a casing U. Both the casing U and the stopcock K are multiply pierced by channels. The stopcock K can be rotated in the casing U in such a manner that in each case defined channels in the stopcock and casing are connected and liquids thus pass from storage vessels via the apparatus shown in a defined manner into the connected isotachophoresis instrument. Storage vessels and the ITP instrument are not shown in the figure, but only indicated by arrows. In Figures ~~A1/A2~~ 4A/4B, the stopcock is rotated so that there is a connection between channel pieces 3, 4 and 5, and between 2 and 6. By this means, channel piece 5 in the interior of the stopcock is filled with sample solution from a storage vessel which is connected to channel 3. In addition, via a storage vessel on channel 2, the channel system of the isotachophoresis instrument is filled with one of the two separation buffers (buffer 1) necessary for ITP.

**On page 13, the first full paragraph has been amended as follows:**

In a second step (Figure ~~B1/B2~~ 4C/4D), the stopcock K is rotated so that the channel connections existing in Figure ~~A1/A2~~ 4A/4B are broken. Instead, a connection is made between channel pieces 1 and 7. In this manner, the channel system situated downstream of the delivery apparatus is filled with a second buffer (buffer 2). In Figure ~~C1/C2~~ 4E/4F,

finally stopcock K is rotated again so that a connection is formed between channel pieces 1, 5 and 2. Channel 2 is filled with buffer 1, channel 5 with the sample solution and channel 1 with buffer 2. In this manner, a sample solution volume defined by the dimensions of channel 5 is embedded between the two buffers necessary for ITP. By applying a voltage, the separation can then be begun.

**In the Drawings:**

Applicant's propose to amend the drawings as shown in red in the attached copy of the drawings to correct the Figure designations.